

Seneganolide, a Novel Antifeeding Mexicanolide from *Khaya senegalensis*

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Seneganolide, a new mexicanolide type rings B,D-seco limonoid was isolated as insect antifeedant together with a known methyl angolensate from the stem bark of *Khaya senegalensis* (Meliaceae). The structure was elucidated on the basis of spectroscopic means and the antifeedant activity was tested by a conventional leaf disk method.

Khaya senegalensis (Desr.) A. Juss. is one of the most popular medicinal plants in the African traditional remedies. The decoction of the bark is extensively used as febrifuge which could be associated with its use as an antimalarial drug.¹ This genus is the main source of African mahogany and is closely related to the South American genus *Swietenia*, which is one of the main source of rings B,D-seco limonoid mexicanolides,² which were biogenetically derived from 1,3-diketodiene lactones of the andirobin-type B,D-seco limonoids like methyl angolensate (**2**) by spontaneous Michael cyclization.³ Some B,D-seco limonoids containing the mexicanolide have been also reported from *K. senegalensis*.³⁻⁵ In a series of our experiments on Meliaceae plants, several types of limonoids were isolated as insect antifeedant from *Trichilia roka*,⁶ *Melia azedarach*⁷ and *M. toosendan*.⁸ In view of our interest in the antifeedant activity of members of the family Meliaceae, we have studied the ether extract of the bark of *K. senegalensis*. Droplet countercurrent chromatography (DCCC) with descending mode of the extract, followed by reversed phase HPLC purification, resulted in the isolation of one novel limonoid belonging to the mexicanolide, named seneganolide (**1**), together with a known rings B,D-seco compound, methyl angolensate (**2**).⁹ In this paper, we report the structure of **1** and its antifeedant activity against the larvae of a Japanese insect pest *Spodoptera littoralis* by a leaf disk method.¹⁰

Seneganolide (**1**) (5 mg),¹¹ mp 276–278 °C (decomp) and $[\alpha]_D^{25} -27^\circ$ (c 0.11, MeOH), was obtained as white needles from acetone/hexane. Its molecular formula C₂₆H₃₀O₈ (12 unsaturations) was determined by HRFAB-MS (m/z 471.2021 [M+1]⁺; $\Delta +0.2$ mmu). The UV maximum at 210 nm (ϵ 5000) and IR absorption at 3580–3300, 1730, 1700, 1637 and 875 cm⁻¹ showed the presence of carbon–carbon double bonds and hydroxyl and carbonyl (keto and ester) groups. From its ¹H and ¹³C NMR data (Table 1) measured in CDCl₃ containing a small amount of CD₃OD owing to the poor solubility of **1** in CDCl₃, it was evident that five of the elements of unsaturation were present as double bonds: two carbon–carbon (as a furan ring), three CO (two as esters and one ketone). Therefore, the molecule is heptacyclic. The NMR data also revealed that compound **1** has three methyls, six methylenes, eight methines (three olefinic), nine quaternary carbons (one olefinic) and one proton due to OH group. Furthermore, the spectra showed the presence of a β -furyl moiety.

After assignment of all protons directly bonded with carbon atoms by the ¹H–¹³C shift-correlated measurement (HMOC spectrum), it was possible to assume from the ¹H–¹H COSY spectrum, decouplings and HMBC spectrum (Figure 2) that **1** was a mexicanolide. The chemical shifts and couplings for protons attached to carbons—17, 5 and 6; 2 and 30; 9, 11 and 12; 14 and 15—also interpreted the presence of the carbon skeleton of **1**.

Especially, the presence of the hemiacetal linkage of C-1 to C-8 in **1** was confirmed from the chemical shifts of C-1 (δ 107.7, s) and C-8 (δ 80.8, s) and from a W-type long range coupling between 30 α and 9 α -H signals. The same acetal linkage of C-1 to C-8 has been observed in some mexicanolides,¹²⁻¹⁴ in which the ring C is present in a skew boat form. A W-type long range coupling between 12 β and 15 β -H signals and NOEs (Figure 3) between 5 β and 11 β -H, 9 α and 14 α -H, 14 α and 18-H (13 α -Me), 15 β and 30 α -H, and 30 α and 2-H signals also elucidated the relative configuration of four chiral centers at C-8, 9, 13 and 14.

The presence of three methyls at 4 α (29), 4 β (28) and 13 α (18) and that the remaining methyl group (19) to be at C-10 in the basic mexicanolide skeleton was changed to a methylene group directly attached to an oxycarbonyl group was confirmed from the HMBC spectrum. Then, the relative stereochemical

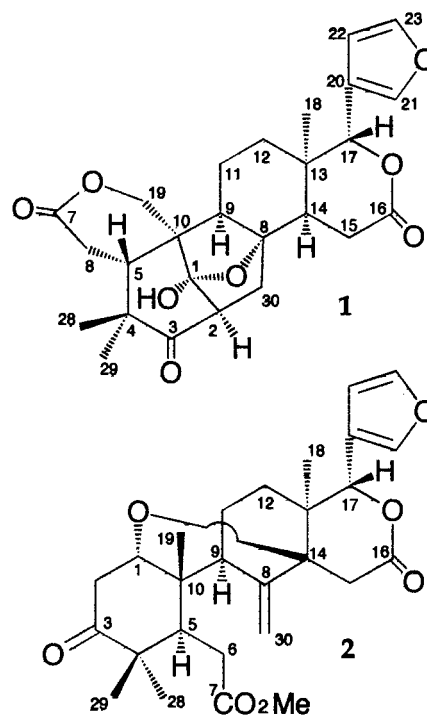


Figure 1. Structure of compounds **1** and **2**.

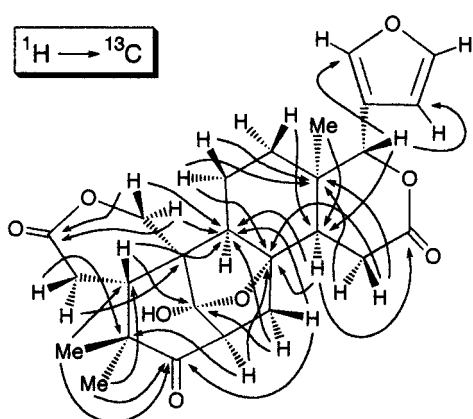


Figure 2. Significant HMBC correlations in 1.

assignment of all of the protons in 1 was established from the NOESY experiment (Figure 3). Although many rings B,D-seco limonoids have been reported, this is, to the best of our knowledge, the first report of occurrence of C-19 oxygenated compound in mexicanolides.

The antifeedant activity of seneganolide (1) and methyl angolensate (2) was tested by a conventional leaf disk method against the third-instar larvae of a Japanese insect pest *Spodoptera littoralis* (Boisduval). Seneganolide (1) was active at 300 ppm, corresponding to a concentration of 6 $\mu\text{g}/\text{cm}^2$, and the compound 2 at 500 ppm. Their potent activities were comparable to those of trichilins⁶ and azedarachins⁷ from meliaceous plants.

Table 1. ^{13}C and ^1H NMR data for seneganolide (1)

atom	δ_{C}	δ_{H}	atom	δ_{C}	δ_{H}
1	107.7 s		15 α	27.8 t	2.80 dd (19.8, 2.2)
2	53.8 d	2.86 dd (13.2, 6.8)	β		2.85 dd (19.8, 7.0)
3	212.1 s		16	170.6 s	
4	44.9 s		17	78.3 d	5.26 s
5	38.7 d	2.30 dd (11.0, 7.0)	18	22.6 q	1.03 s
6 α	29.6 t	2.68 dd (14.9, 11.0)	19 α	73.8 t	4.48 d (11.7)
β		2.56 dd (14.9, 7.0)	β		4.21 d (11.7)
7	173.3 s		20	120.7s	
8	80.8 s		21	140.9 d	7.38 br s
9	61.5 d	2.02 br d (9.7)	22	109.7 d	6.31 br d (1.5)
10	46.9 s		23	143.2 d	7.40 t (1.5)
11 α	21.1 t	1.63 m	28	24.1 q	1.19 s
β		1.41 m	29	19.6 q	1.31 s
12 α	35.1 t	1.43 br t (12.2)	30 α	31.9 t	2.23 td (13.2, 1.9)
β		1.78 dd (11.0, 2.7)	β		1.85 dd (13.2, 6.8)
13	35.5 s		OH		1.88 br
14	44.9 d	2.27 dd (7.0, 2.1)			

Measured in CDCl_3 containing a small amount of CD_3OD at 40°C at 125 and 500 MHz. J values (in Hz) are presented in parentheses.

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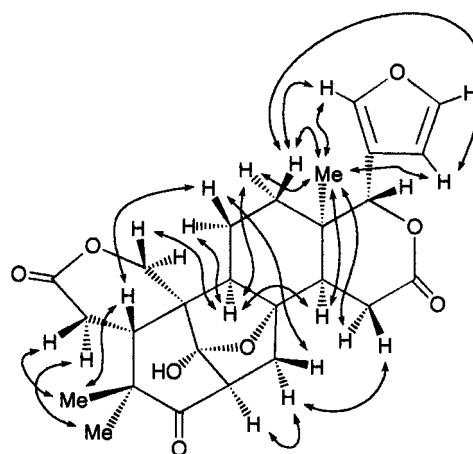


Figure 3. Selected NOE correlations in 1.

References and Notes

- J. M. Watt and M. G. Breyer-Brondwijk, "The Medicinal and Poisonous Plants of Southern and Eastern Africa," 2nd ed., London Livingstone, London (1962).
- D. A. Okorie and D. A. H. Taylor, *Phytochemistry*, **10**, 469 (1971).
- D. A. H. Taylor, "The Chemistry of the Limonoids from Meliaceae," in "Progress in the Chemistry of Organic Natural Products," ed. by W. Herz, H. Grisebach, and G. W. Kirby, Springer, New York (1984), p. 1.
- L. R. V. Olmo, M. F. da Silva, E. Rodrigues Fo, P. C. Vieira, J. B. Fernandes, A. J. Marsaioli, A. L. Pinheiro, and E. F. Vilela, *Phytochemistry*, **42**, 831 (1996).
- L. R. V. Olmo, M. F. da Silva, E. Rodrigues Fo, P. C. Vieira, J. B. Fernandes, A. L. Pinheiro, and E. F. Vilela, *Phytochemistry*, **44**, 1157 (1997).
- M. Nakatani and K. Nakanishi, *Heterocycles*, **36**, 725 (1993) and references cited therein.
- M. Nakatani, "Insect Antifeeding Limonoids from the Chinaberry Tree *Melia azedarach* Linn. and Related Compounds," in "The Biology-Chemistry Interface," ed. R. Cooper and J. K. Snyder, Marcel Dekker, Inc., New York (1999), p. 1.
- M. Nakatani, *Heterocycles*, **50**, 595 (1999).
- G. A. Adesida, E. K. Adesogan, and D. A. H. Taylor, *J. Chem. Soc., Chem. Commun.*, **1967**, 790.
- K. Wada and K. Munakata, *J. Agr. Food Chem.*, **16**, 471 (1968).
- Compounds 1 (5 mg) and 2 (2.4 mg) were finally isolated by HPLC with 55 and 45% H_2O -MeOH solvent systems from the extract (3.9 g), respectively.
- J. D. Connolly, M. MacLellan, D. A. Okorie, and D. A. H. Taylor, *J. Chem. Soc., Perkin 1*, **1976**, 1993.
- D. A. H. Taylor, *Phytochemistry*, **22**, 1297 (1983).
- W. M. Daniewski, M. Gumulka, W. Danikiewics, P. Gluzinski, J. Krajewski, J. Sitkowski, E. Bloszyk, U. Jacobsson, T. Norin, and F. Szafranski, *Phytochemistry*, **36**, 1001 (1994).